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Jiayue Guo – University of Alabama

Peilong Li – Cornell University

Lingyan Kong – University of Alabama

Baojun Xu – Beijing Normal University

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or send request to lingyan.kong@ua.edu

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Microencapsulation of Curcumin by Spray Drying and Freeze Drying

Jiayue Guo ^a, Peilong Li ^b, Lingyan Kong ^{a,*}, Baojun Xu ^{c,*}

^a Department of Human Nutrition & Hospitality Management, The University of Alabama, Tuscaloosa, AL 35487, United States

^b Department of Food Science, Cornell University, Ithaca, NY 14853, United States

^c Food Science and Technology Program, Beijing Normal University– Hong Kong Baptist University United International College, Zhuhai, Guangdong 519087, China

*Corresponding authors.

Address:

2000 Jintong Road, Tangjiawan, Zhuhai, Guangdong 519087, China. (B. Xu)

482 Russell Hall, 504 University Blvd, Tuscaloosa, AL 35487, USA (L. Kong)

Email:

baojunxu@uic.edu.hk (B. Xu)

lkong@ches.ua.edu (L. Kong)

26 **Abstract**

27 Curcumin is a natural pigment with health benefits and potential uses in food and
28 pharmaceutical industries, but its application is limited by its insolubility and instability. This
29 study was to examine the effect of spray drying and freeze drying methods, as well as 12
30 combinations of ternary-composite wall materials, on microencapsulation efficiency (MEE) ,
31 physicochemical properties (including particle size distribution, morphology, moisture content,
32 and color values), and stability of curcumin against environmental stresses and in a model
33 beverage. Curcumin encapsulated by freeze drying presented higher MEE, whereas spray drying
34 produced microparticles had smaller particle sizes, smoother granule surface, and more regular
35 shapes. Though to different extent, curcumin encapsulated by both methods exhibited improved
36 stability against heat and acidity, and improved stability in a carbonated beverage. This study
37 provides practical information on the selection of wall materials and encapsulation methods for
38 microencapsulating curcumin for various applications.

39

40 **Keywords:** Curcumin; microencapsulation; spray drying; freeze drying; wall materials;
41 microencapsulation efficiency

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44 **1. Introduction**

45 Curcumin is a yellow-orange pigment produced by turmeric (*Curcuma longa*), a member of
46 the ginger family. As a natural phytochemical, curcumin exhibits a wide range of bioactivities,
47 e.g., antioxidative, and anti-inflammatory properties, marking its significance in clinical
48 utilization (Hewlings & Kalman, 2017; Wilken, Veena, Wang, & Srivatsan, 2011). However,
49 free curcumin has little confirmed uses or practical applications in both food processing and
50 medical fields, as it is insoluble in aqueous solutions, susceptible to heat, light, and extreme
51 acidity, meaning that it can be easily degraded under environmental stresses (Liu et al., 2017;
52 Nelson et al., 2017; Wang, Lu, Lv, & Bie, 2009).

53 Microencapsulation is a widely used technology to improve the stability and bioavailability
54 of bioactive compounds (Gharsallaoui, Roudaut, Chambin, Voilley, & Saurel, 2007).
55 Microcapsules or microparticles are the product of microencapsulation and contains the
56 bioactives in the core surrounded by a coating of wall materials. The wall provides a physical
57 barrier that prevents molecular diffusion and chemical reactions so as to improve the stability of
58 encapsulated compounds (Taguchi, Saito, Uchida, & Tanaka, 2016). Various microencapsulation
59 techniques have been invented, such as spray drying, freeze drying, extrusion, and centrifugal
60 extrusion among others. Among them, spray drying is the most commonly used technique
61 because of its cost-efficiency, flexibility, and continuous operation (Gouin, 2004; Jafari,
62 Assadpoor, He, & Bhandari, 2008; Ré, 2006). Freeze drying is another popular method,
63 especially for the more sensitive and expensive bioactive compounds. Unlike spray drying,
64 freeze drying uses a much lower temperature and operates in the absence of oxygen. Numerous
65 studies employed spray drying and freeze drying techniques to encapsulate curcumin (Cano-
66 Higuera, Malacrida, & Telis, 2015; Soudaleff et al., 2013; Zuanon, Malacrida, & Telis, 2013),

67 yet, few research compared various combinations of ternary composite wall materials with both
68 microencapsulation methods.

69 The objective of this study was to examine the effect of both spray drying and freeze drying
70 methods, as well as 12 different combinations of ternary composite wall materials, on
71 microencapsulation efficiency and stability of curcumin against environmental stresses.
72 Additionally, this study also examined the application of microencapsulated curcumin in a
73 carbonated beverage, which would provide reference information for the food industry.

74 **2. Materials and methods**

75 *2.1. Materials:*

76 Curcumin (90-95% purity) was provided by Yafuxingyuan Food Industry Co., Ltd. (Zhuhai,
77 China). Modified corn starch (MS, water soluble corn starch) and maltodextrin (MD) with
78 dextrose equivalent of 20 were obtained from Zhiyou Technology Co., Ltd. (Changchun, China).
79 Inulin (IN), gelatin (GE), tamarind gum (TG) were obtained from Beneo Orafit SA (Belgium),
80 Amin Biological Muslim Gelatin Co., Ltd. (Lanzhou, China), and BLX Food Materials Co., Ltd.
81 (Zhengzhou, China), respectively. β -Cyclodextrin (CD) and pectin (PE) were obtained from
82 Huayue Chemical Products Co., Ltd. (Zhengzhou, China). All the chemical reagents used were
83 of analytical grade.

84 *2.2. Formulation and preparation of emulsion*

85 To prepare the emulsions for spray drying and freeze drying, 12 combinations of wall
86 materials were formulated according to **Table 1**. Wall materials (30 g) were dissolved in distilled
87 water and mixed at 1,008 g until complete dissolution. After setting still overnight, 1 g of
88 curcumin powder in 40 mL of absolute ethanol was poured into the wall materials solution and
89 mixed using a high-speed homogenizer (T25 digital Ultra Turrax, IKA, Germany) at 28,672 g for

90 10 min in cold water bath to reach complete emulsification. A small amount of distilled water
91 was used to rinse the beaker and the curcumin residue to the emulsion, and the emulsion was
92 made up to 100 mL.

93 *2.3. Microencapsulation by spray drying and freeze drying*

94 Spray drying was carried out using a lab-scale spray drier (Laiheng L-217, Beijing, China),
95 equipped with a spray-drying chamber, a high-speed atomizer, a cyclone separator, a hot air
96 blower, an air compressor with a maximum exhaust pressure of 0.75 MPa, and a spray nozzle of
97 0.7 mm. The homogenized emulsion was fed into the chamber with inlet temperature of 190 °C
98 and feed rate of 600 mL/h, atomized by the hot air (blow rate of 28 m³/s), and the spray-dried
99 microparticles were collected in a glass collector.

100 For freeze drying, core-wall emulsions were frozen at -80 °C for more than 12 h, and then
101 placed in a Labconco FreeZone 1 freeze drier (Kansas City, MO, USA) at -45 °C, lower than 10
102 Pa for 2 -3 days. The freeze dried samples were then ground into fine powder with a mortar and
103 pestle.

104 *2.4. Determination of microencapsulation efficiency (MEE)*

105 The amount of curcumin was quantified by spectrophotometric measurement at 425 nm
106 using a spectrophotometer (Jingje, Shanghai, China). The calibration curve ($R^2=0.9998$) was
107 established using a series of curcumin solutions in ethanol ranging from 1 to 5mg/L. The
108 microencapsulation efficiency (MEE) of curcumin microparticles was calculated according to
109 Eq. 1.

$$110 \quad \text{MEE} = (\text{Total curcumin} - \text{superficial curcumin}) / \text{Total curcumin} \times 100\% \quad (\text{Eq. 1})$$

111 Where superficial curcumin is the amount of curcumin that was partitioned into the ethanol
112 phase when the microparticles were mixed into absolute ethanol, and total curcumin is the
113 amount of curcumin extracted into ethanol after dissolving the microparticles in water.

114 *2.5. Morphological analyses*

115 The particle size distribution of curcumin microparticles was analyzed by a laser diffraction
116 particle size analyzer (LA-920, Horiba Ltd., Tokyo, Japan) in the wet mode, with measuring
117 range from 0.02 to 2000 μm . Powder samples were suspended in isopropyl alcohol, sonicated
118 with a microtip probe, and particle size distribution was determined after successive readings
119 became constant. For the evaluation of surface morphology, powder samples were deposited on
120 carbon double-sided adhesive tape mounted on stubs, which were observed using a scanning
121 electron microscope (SEM, JSM-6390A, JEOL, Tokyo, Japan) with an accelerating voltage of 15
122 keV.

123 *2.6. Moisture content analysis*

124 The moisture content of microparticle samples was determined using a direct drying method
125 based on the AOAC method with slight modifications (AOAC, 2005). In detail, dry and clean
126 aluminum pans were weighed (m_0) and loaded with 0.5 g of microparticle powder. Samples were
127 then dried for 4 h at 70 °C in vacuum oven (Mettler Co., Shanghai, China) for complete
128 dehydration. Aluminum pans with samples after drying were then weighed again (m_1) until the
129 dried weight stayed the same. Moisture content of microparticles were calculated as $[1 - (m_1 - m_0$
130 $/ 0.5)] \times 100\%$.

131 *2.7. Color analysis*

132 Color values of curcumin microparticle samples were measured using a HunterLab D25
133 colorimeter (Hunter Lab Associates Inc, Reston, VA, USA). The colorimeter was calibrated with

134 a CR-A43 calibration white plate. Values including L^* (lightness), a^* (redness and greenness),
135 and b^* (yellowness and blueness) were recorded. In detail, L^* stands for the lightness ranging
136 from 0 (black) to 100 (white); a^* indicates redness with “+” and greenness with “-”; while
137 b^* indicates yellowness with “+” and blueness with “-”.

138 *2.8. Stability tests*

139 The heat stability of curcumin was measured as the retention (%) of curcumin after the
140 microparticle suspensions (0.2%, w/v) were heated at 90 °C for 20 min. The acid stability of
141 curcumin was determined by its retention (%) after the microparticle suspensions (0.2%, w/v)
142 were adjusted to pH of 1 by adding hydrochloric acid. Stability of free curcumin was measured
143 using its ethanol solution. The amount of curcumin was determined by the same
144 spectrophotometric method as described in 2.4.

145 *2.9. Stability of microparticles in carbonated beverage*

146 The microparticles were mixed into a transparent carbonated beverage, Sprite (Coca-Cola,
147 Atlanta, GA, USA), to reach a 0.05% (w/v) dispersion. The beverage was then set still at room
148 temperature for 48 h for the quantitative evaluation of stability (% retention), and visual
149 evaluation of clarity and sediment formation.

150 *2.10. Statistical analysis*

151 All the experiments were conducted in triplicates, and the results were expressed as mean
152 value \pm standard deviation and analyzed by one-way analysis of variance (ANOVA) with
153 Duncan’s Multiple Range Test to determine the significant differences among the means ($p <$
154 0.05), using the SPSS 17.0 software.

155 **3. Results and discussion**

156 *3.1. Microencapsulation efficiency (MEE) of curcumin*

157 The MEE of curcumin in spray dried and freeze dried microparticles are summarized in
158 **Table 2**. In general, freeze dried microparticles showed higher MEE than the spray dried
159 samples, except that freeze dried formula B, J, and L resulted in lower MEE. The MEE of most
160 freeze dried microparticles was above 70 % except for the two formula using inulin (IN) and β -
161 cyclodextrin (CD) (J and L). Formula I, which used IN, maltodextrin (MD), and tamarind gum
162 (TG) as wall materials, resulted in the highest MEE among all freeze dried and spray dried
163 microparticles, i.e., 89.44% and 82.50%, respectively. The higher retention of active compounds
164 by freeze drying than by spray drying was also reported by other researchers (Ballesteros,
165 Ramirez, Orrego, Teixeira, & Mussatto, 2017; Murali, Kar, Mohapatra, & Kalia, 2015; Ramírez,
166 Giraldo, & Orrego, 2015). The reason can be attributed to the milder processing conditions of
167 freeze drying (low temperature and absence of oxygen) compared with spray drying (high
168 temperature and presence of oxygen) (Cano-Higueta et al., 2015; Chen, Zhong, Wen,
169 McGillivray, & Quek, 2013). An elevated temperature (190 °C in spray drying) and air flow
170 providing oxygen accelerated the degradation of curcumin (Aniesrani Delfiya, Thangavel,
171 Natarajan, Kasthuri, & Kailappan, 2015). Therefore, for sensitive materials like curcumin, freeze
172 drying could be a better choice for its microencapsulation. As for different formulas, the lowest
173 MEE of curcumin (54%) in freeze dried samples was resulted from formula L (IN, CD, and TG).
174 TG is a group of structural heteropolysaccharides from tamarind plants, which is composed of a
175 1,4-linked β -D glucan main chain and α -D xylan side chains (Keonakhone et al., 2017).
176 Although being widely used in industrial applications, such as glue, thickener, and paper
177 adhesive manufacturing, TG has many drawbacks such as low solubility in cold water and

178 biodegradability. Such drawbacks limited its efficiency in forming microparticles with high MEE
179 as one of the wall materials. Similar results have been reported by Keonakhone et al. (2017),
180 indicating that TG was not completely dissolved in water and presented a higher viscosity that
181 caused droplet aggregation, thus resulting a lower EE value of microparticles produced. In
182 addition, the only difference between formula L and I was the auxiliary material used: CD vs.
183 MD, but the MEE of formula I (IN, MD, and TG) was much higher than that of formula L ($p <$
184 0.05). Similar results were also found among microparticles embedded with IN as structural
185 material: high-MEE formulas G and H vs. low-MEE formulas J and K in freeze drying, and H
186 and I vs. K and L in spray drying. Therefore, based on the above results, it can be inferred that
187 IN worked better with MD than CD for trapping curcumin into the microparticles, with higher
188 MEEs obtained. Another study investigating encapsulating carbohydrates also indicated that the
189 mixture of octenylsuccinylated starch, inulin, and MD was the best combination of encapsulating
190 wall material, which produced jussara pulp microparticles of desired properties (Lacerda et al.,
191 2016).

192 *3.2. Particle size and morphology of curcumin microparticles*

193 The particle size distribution patterns of curcumin microparticles are presented in **Figure 1**.
194 Spray drying tended to create smaller particles than freeze drying; most spray dried
195 microparticles distributed around 10 μm , while all the freeze dried particles distributed around
196 100 μm . Nearly half of spray dried samples presented unimodal size distribution, while almost
197 all freeze dried microparticles exhibited bimodal distribution pattern. Spray drying of formulas A
198 and C exhibited the finest and most uniform microparticles, with 99% of particles having
199 diameter below 100 μm . The difference in particle size distribution, especially affected by the
200 two methods, was because of the materials and processes involved. Spray drying applies

201 powerful atomization, whereas in freeze drying, the final particle size only depended on the
202 grinding procedure instead of the drying process (Cano-Higueta et al., 2015).

203 The morphological structures of microparticles examined by SEM are presented in **Figure 2**.
204 In addition to their difference in particle size, which is in agreement with the particle size
205 distribution patterns of **Figure 1**, microparticles created by the two drying methods resulted in
206 very different morphological characteristics. Spray dried microparticles were more spherical and
207 regular in shape, and their surface appeared relatively smoother than the freeze dried ones, which
208 exhibited more cracks or fractures. However, some spray dried microparticles tended to fuse
209 together (B, F, I, and K), which may be due to the viscosity, hygroscopicity, and concentration of
210 the wall materials, as well as their film forming capacity.

211 *3.3. Physical characteristics of curcumin microparticles*

212 Physical properties of curcumin microparticles, including moisture content, color, and
213 pigment value, are shown in **Table S1** in Supplementary Materials. The moisture content of
214 curcumin microparticles ranged from 1.9% to 10.1 % (spray dried) and from 1.4% to 8.6 %
215 (freeze dried), while most treatments resulted in moisture content below or around 5 %. This
216 range agreed with Cano-Higueta et al. (2015), in which the moisture content of curcumin
217 microparticles was all below 5% using different wall material compositions by both freeze
218 drying and spray drying. Furthermore, the two drying methods created significant difference in
219 moisture content. Specifically, freeze dried microparticles exhibited lower moisture content in
220 most cases excepting formula G, indicating more dehydration through freeze drying. Group J (IN
221 + CD + GE) exhibited a very high moisture content by both drying methods (8% - 10%). By
222 spray drying, microparticles embedded with formulas A, B, and F also presented high moisture

223 contents (> 4%), which served as one reasonable explanation why the particles stuck or merged
224 together as observed from the SEM images.

225 Color values reflected the lightness and saturation degree of curcumin microparticle
226 products (**Table S1** in Supplementary Material). All curcumin microparticles presented high
227 lightness with L^* ranging from 79.58 to 93.62. Most of the a^* values were slightly above 0,
228 indicating a small tendency to redness, whereas the high b^* values above 0 indicated significant
229 yellow tendency due to their curcumin content. Spray dried samples embedded with MS and CD
230 (E and F) even possessed b^* values greater than 90. The b^* value could be an important indicator
231 for the quality of curcumin microparticles. The spray dried and freeze dried microparticles
232 exhibited b^* values ranging from 75.12 (H) to 91.48 (E), and from 58.29 (I) to 81.77 (G),
233 respectively. Therefore, the overall yellowness of spray dried microparticles were more than that
234 of freeze dried samples, although freeze dried microparticles showed generally higher MEE
235 (**Table 2**). It was possible that higher surficial curcumin contributed more to the yellowness than
236 curcumin trapped in the core and coated with white wall materials. In particular, the
237 microparticles made with formula I by freeze drying presented the lowest b^* value (58.29), yet,
238 gave the highest MEE (89.44%, **Table 2**).

239 *3.4. Stability of curcumin microparticles against environmental stresses*

240 The stability of microencapsulated curcumin against high temperature and acidic conditions,
241 expressed as percent retention, is presented in **Table 2**. The retention rates of free curcumin
242 under the same environmental stresses were 60.8% and 79.9%, respectively. Both were
243 significantly lower than any of the microencapsulated curcumin. The results were consistent with
244 the current literature. A recent review on curcumin encapsulation reported that the chemical
245 stability of curcumin is enhanced by encapsulating it in microemulsions (Jiang, Liao, &

246 Charcosset, 2020). Yet, there was no simple trend that spray dried microparticles demonstrated
247 higher stability than freeze dried samples, or vice versa. The highest thermal stability was
248 achieved using formula D by spray drying and formula A by freeze drying. The highest acid
249 stability was seen in spray dried formula D & F, and in freeze dried formula F. The wall
250 materials used and morphological characteristics should all affect the heat and acid resistance of
251 the microparticles. In several wall material combinations, modified corn starch as the structural
252 material tended to provide better protection against environmental stresses than inulin. For
253 instance, formula I with the highest MEE did not show high thermal and acid stability. As most
254 food products present pH values higher than 2, it was ensured that curcumin microparticle could
255 be applied to highly acidic foods with acceptable resistance to acid even when the pH value was
256 1.

257 *3.5. Application of curcumin microparticles in carbonated beverage*

258 Most carbonated beverages, e.g., Sprite (pH=3.3) used in this study, are slightly acidic with
259 pH in the range of 3 to 4, due to dissolved carbon dioxide gas and added organic acids. All the
260 curcumin microparticles were soluble or dispersible in the beverage samples. The retention rates
261 of curcumin after 2 days in Sprite beverage are shown in **Table 2**. Different extents of sediment
262 formation occurred among samples with different wall materials (**Table S2** in Supplementary
263 Material) and the retention of curcumin ranged between 55% and 97%. Among them, sample
264 groups E, F, G, H, K performed well in carbonated beverage for the clarity and absence of
265 sedimentation with no matter which drying method was applied. Formulas A, G, H, and J
266 produced by spray drying, as well as groups G and J by freeze drying exhibited high retention of
267 curcumin after the 2-day settling, which were above 90%. It seemed that inulin performed better
268 than modified corn starch as a structural material for the application in carbonated beverage. In

269 addition to its stabilizing effect in beverage, inulin could also serve as a supplement of dietary
270 fiber, and could be welcomed by customers pursuing healthy eating, which marked its potential
271 application in the beverage industry for its use in microencapsulation.

272 **4. Conclusion**

273 Curcumin microparticles formulated with 12 combinations of wall materials were produced
274 by spray drying and freeze drying. Microencapsulation efficiency of freeze drying was generally
275 higher than that of spray drying. Spray drying presented finer, and more regular and spherical
276 particles. The average moisture content of microparticles was around or below 5% in most
277 samples and all of the samples exhibited similar lightness. Spray dried microparticles were more
278 yellow than freeze dried ones, possibly owing to more superficial curcumin and less
279 encapsulated curcumin. Microencapsulated curcumin showed enhanced stability against high
280 temperature and acidic conditions than free curcumin. While formulas with modified starch as
281 structural material presented higher thermal and acid stability than those with inulin. Inulin-
282 based formulas performed better in carbonated beverage, making it a good choice as coating
283 material for its encapsulating performance and health beneficial effects. This study provides
284 invaluable reference on the selection of wall materials and processes for future studies on the
285 microencapsulation of curcumin and other bioactive compounds.

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